



Short Communication

Phenotypic plasticity and spatial distribution of *Simulium pertinax* larvae from the Tijuca National Park

Plasticidade fenotípica e distribuição espacial de larvas de *Simulium pertinax* do Parque Nacional da Tijuca

Marlon José Ribeiro Pinto¹, Agatha Alves da Silva¹, Bárbara Alves Victér¹, Milena Cavalcanti Pinto Lopes¹, Tainá Maria Miranda Souza-Martins^{1,2}, Ronaldo Figueiró¹

AUTHOR AFILIATIONS

1 – Universidade do Estado do Rio de Janeiro, Faculdade de Ciências Biológicas e Saúde, Departamento de Biologia, Laboratório de Meio Ambiente e Saúde (LabMAS)
2 – Universidade Federal do Rio de Janeiro, Museu Nacional, Laboratório de Dipteros Aquáticos

ORCIDS AND CONTACT

Marlon José Ribeiro Pinto
Orcid: 0009-0002-9731-4462
ribeiro.mjr@gmail.com
Agatha Alves da Silva
Orcid: 0009-0002-8300-7306
agatharjalvess@gmail.com
Barbara Alves Victér
Orcid: 0009-0007-0972-870X
barbaravictér.bio@gmail.com
Milena Cavalcanti Ponto Lopes
Orcid: 0009-0004-11604864
mlenenacavalcanti13@icloud.com
Tainá Maria Miranda Souza-Martins
Orcid: 0000-0002-1285-7031
tainamiranda.mms@gmail.com
Ronaldo Figueiró
Orcid: 0000-0003-0762-1312
ronaldofigueiro@gmail.com

ABSTRACT

Black fly larvae are filter feeding organisms that inhabit lotic systems and are considered key-organisms for their role as trophic link converting Fine Particles Organic Matter (FPOM) into Compact Particles Organic Matter (CPOM) making it available to a broader range of microorganisms. Literature suggests that black fly larvae may present phenotypic plasticity due to a wide range of physical factors such as water velocity, as well as food availability. The present study investigates the variation of morphological characteristics of *Simulium pertinax* larvae among different microhabitats in the Tijuca National Park, finding significant correlations among proleg length, anal disk and cephalic capsule sizes and the water velocity.

Keywords: Black flies. Bioindicator, Biomonitoring.

RESUMO

As larvas de simulídeos são organismos filtradores que habitam sistemas lóticos e são considerados organismos-chave por seu papel como elo trófico, convertendo matéria orgânica de partículas finas (FPOM) em matéria orgânica de partículas compactas (CPOM), tornando-a disponível para uma gama mais ampla de microrganismos. A literatura sugere que larvas de simulídeos podem apresentar plasticidade fenotípica devido a uma ampla gama de fatores físicos, como velocidade da água, bem como disponibilidade de alimento. O presente estudo investiga a variação das características morfológicas de larvas de *Simulium pertinax* entre diferentes microhabitats no Parque Nacional da Tijuca, encontrando correlações significativas entre o comprimento da perna, o tamanho do disco anal, da cápsula cefálica e a velocidade da água.

Palavras-chave: Borrachudos, Bioindicador, Biomonitoramento.

Members of the Simuliidae family, commonly known as black flies or "piums" in Brazil, are dipterans with a worldwide distribution, found everywhere except for the Antarctic region, deserts, and islands devoid of streams (CROSSKEY, 1990). The larvae are aquatic and filter-feeding, consuming fine organic particles dissolved and suspended in water (ALENCAR et al. 2001). They are commonly found in rivers with high dissolved oxygen content and varying levels of organic matter (STRIEDER et al. 2002). Currently, there are 2,407 valid species of Simuliidae, with 97 species recorded in Brazil (ADLER, 2024). Among the Simuliidae species found in Brazil, *Simulium pertinax* Kollar is considered the main anthropophilic species in the southeastern region (ARAÚJO-COUTINHO et al. 1988), where it is found in large densities and becomes an annoyance to humans and domestic animals, affecting tourism and agriculture activities (ARAÚJO-COUTINHO & LACEY, 1990). Since the larvae of Simuliidae are rheophilic, meaning they depend on currents (ALENCAR et al. 2001), larvae of different species can be found together, depending on different current speed ranges, and water variability can influence the heterogeneity of Simuliidae habitats, directly correlating with species diversity (FIGUEIRÓ et al. 2008; 2012).

Environmental factors, such as water speed, combined with ontogenetic factors, can influence

the growth rate of an organism (BRIERS et al. 2004), impacting the phenotypic plasticity, which is the ability of organisms to alter their physiology or morphology in response to environmental conditions (NYLIN; GOTTHARD, 1998). The aim of this study is to investigate the phenotypic plasticity in *Simulium pertinax*, seeking to identify correlations between its morphology and the microhabitat it occupies.

The study area was the Tijuca National Park, located in the city of Rio de Janeiro, in the mountains of the Tijuca Massif, between the parallels 22°55'S and 23°00'S and the meridians 43°11'W and 43°19'W, a conservation unit that protects fragments of the Atlantic Forest with high biological diversity and good conservation status (ICMBIO, 2008). Collections were made in five sections of streams within the protected areas of Tijuca National Park, and substrates containing Simuliidae larvae were collected from a total of fifteen 30x30 cm quadrats randomly distributed between the stream margins and the center of each site. The average velocity, the dominant substrate type (current leaf litter), and the depth were recorded for each quadrat. Water velocity measurements were taken using the "Head Rod" method (WILM & STOREY, 1944). The larvae were stored in 70% ethanol, and later subjected to morphotyping and species identification using the key by Gil-Azevedo et al. (2005). Final instar specimens, with mature histoblasts, were dissected and mounted between a slide and

coverslip following the method of Calvão & Maia-Herzog (2003). Fourteen *S. pertinax* larvae were selected, and their images were captured using a Motic SMZ140 stereoscopic microscope coupled with a Moticam 5.0 MP image capture device. The obtained images were then measured using the CMEIAS Image Tool software. The morphological measurements were each subjected to linear regression, with water velocity as the independent variable. After performing the linear regression analysis (Figure 1 to 3), a negative correlation was found between the head capsule ($p=0.0385$), proleg ($p=0.0154$), and anal disc ($p=0.0145$) with water velocity, indicating that these structures tended to be smaller at higher water velocities. Larvae with smaller head capsules, proleg, and anal discs were more frequent in faster currents.

Figure 1: Linear regression of current velocity versus anal disc.

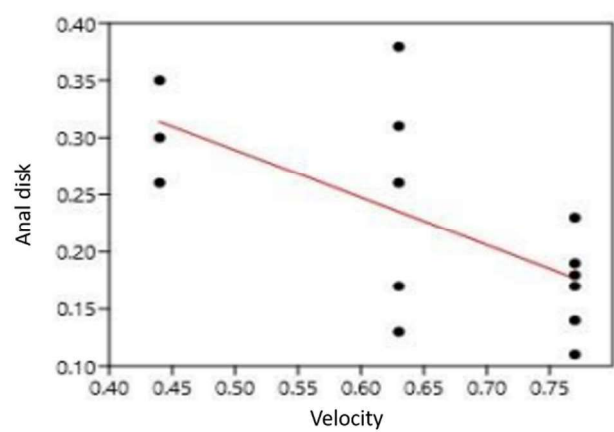


Figure 2: Linear regression of current velocity versus head capsule.

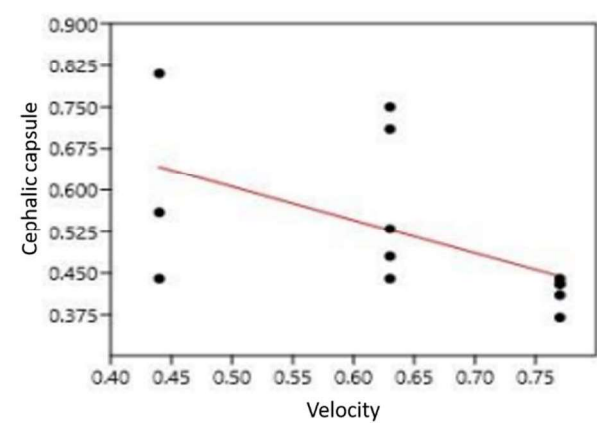
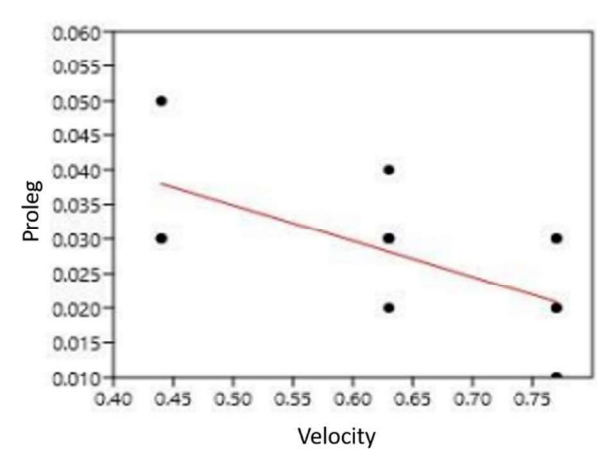


Figure 3: Linear regression of current velocity versus proleg.



These results contrast with those of Figueiró et al. (2015) for the species *Simulium subpallidum* Lutz 1909, where the authors observed a tendency for the structures of the proleg and anal disc to increase at higher current velocities. Such divergence in patterns may be due to species-specific characteristics or even differences between the rivers of the Cerrado biome and those of the Atlantic Forest biome.

Another point is that competition between species may be a factor that explains distribution depending on water speed because larger larvae may competitively exclude smaller larvae from the best locations; thus, smaller larvae are often forced to disperse in search of sufficiently fast flow (HEMPHIL, 1988; 1991). It can be assumed, therefore, that larvae with smaller morphological structures of head capsule, anal disc, and proleg are better adapted to higher current velocities. Studies in the literature suggest that smaller larval structures can reduce the energy cost of resisting the current (ZHANG & MALMQVIST, 1996, 1997; ZHANG, 2000; PALMER & CRAIG, 2000). It can be concluded that phenotypic plasticity was observed in *Simulium pertinax* larvae, with variations in the structures of the head capsule, anal disc, and proleg. The patterns found in this study differ from similar studies in other Brazilian biomes with other species, which may be related to the specificities of each species or different selective pressures.

ACKNOWLEDGEMENTS

The authors would like to thank Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ) for the financial support.

REFERENCES

- ADLER, P. H 2024.. World Black Flies (Diptera: Simuliidae): a comprehensive revision of the taxonomic and geographical inventory. Disponível em: <https://biomia.sites.clemson.edu/pdfs/blackflyinventory.pdf>. Acesso em: 19 dez. 2024.
- ALENCAR, Y. B.; LUDWIG, T. A. V.; SOARES, C. C. & HAMADA, N. 2001. Stomach content analyses of *Simulium perflavum* Roubaud 1906 (Diptera: Simuliidae) larvae from streams in Central Amazônia, Brazil. *Memórias do Instituto Oswaldo Cruz*, 96: 561-576.
- ARAÚJO-COUTINHO, A.; MAIA-HERZOG, M.; SOUZA, B. C. 1988. Levantamento das espécies do gênero *Simulium* Latreille (Diptera, Simuliidae) no litoral norte do Estado de São

Paulo. Revista Brasileira de Entomologia, 2 (1):

11-17.

ARAÚJO-COUTINHO, C. J. P. C.; LACEY, L.

1990. Controle de Simulídeos com concentrado emulsionável de *Bacillus thuringiensis*. Boletín de la Oficina Sanitaria Panamericana, 108: 213-219.

BRIERS, R. A.; GEE, J. H. R.; CARISS, H. M.; GEOGHEGAN, R. 2004. Inter-population dispersal by adult stoneflies detected by stable isotope enrichment. *Freshwater Biology*, 49: 425-431.

CALVÃO-BRITO, R. H.; MAIA-HERZOG, M. 2003. Modificação na técnica para montagem de simulídeos (Insecta, Diptera) em lâmina/lamínula. *Revista Brasileira de Zoologia*, 20 (4): 773-774.

CROSSKEY, R. W. 1990. First update to the taxonomic and geographical inventory of world blackflies (Diptera: Simuliidae). Londres: The Natural History Museum.

FIGUEIRÓ, R.; NASCIMENTO, E. S.; GIL-AZEVEDO, L. H.; MAIA-HERZOG, M.; MONTEIRO, R. F. 2008. Local distribution of blackfly (Diptera, Simuliidae) larvae in two adjacent streams: the role of water current velocity in the diversity of blackfly larvae. *Revista Brasileira de Entomologia*, 52:452-454.

FIGUEIRÓ, R.; BERTAZO, K. 2012. Spatial Distribution of Black Fly (Diptera: Simuliidae) Immatures in a Water Current Velocity Gradient in Aracruz/ES, Brazil. *Revista de Ciências da Vida*, 32 (2): 91-101.

GIL-AZEVEDO, L. H.; FERREIRA, J. N.; MAIA-HERZOG, M. 2005. Chave de identificação de pupas de Simuliidae (Diptera) do Sudeste do Brasil. *Revista Brasileira de Zoologia*, 22(3): 742-752.

HEMPHILL, N. 1988. Competition between two stream dwelling filter-feeders, *Hydropsyche oslari* and *Simulium virgatum*. *Oecologia*, 77: 73–80.

HEMPHILL, N. 1991. Disturbance and Variation in Competition Between Two Stream Insects. *Ecology*, 72:864-872.

HEMBOLD, R. 1965. Mapa geológico do Estado do Guanabara; 3 folhas. Rio de Janeiro: DNPM.

ICMBio. 2008. Plano de manejo: Parque Nacional da Tijuca. Brasília. Disponível em: <http://www.icmbio.gov.br/parnatijuca/plano-de-manejo>. Acesso em: 19 dez. 2024.

NYLIN, S.; GOTTHARD, K. 1998. Plasticity in life-history traits. *Annual Review of Entomology*, 43: 63-83.

PALMER, R. W.; CRAIG, D. A. 2000. An ecological classification of primary labral fans of filter-feeding black fly (Diptera: Simuliidae) larvae. *Canadian Journal of Zoology*, 78 (2): 199-218.

STRIEDER, M. N.; SANTOS, J. E.; PÊS, A. M. O. 2002. Diversidade e distribuição de Simuliidae

(Diptera, Nematocera) no gradiente longitudinal da bacia do rio dos Sinos, no Rio Grande do Sul, Brasil. *Entomology y Vectores*, 9 (4):527-540.

WILM, H. G.; STOREY, H. C. 1944. Velocity-head rod calibrated for measuring streamflow. *Civil Engineer*, 14: 475-476.

ZHANG, Y. 2000. Effects of fan morphology and habitat on feeding performance of blackfly larvae. *Archiv für Hydrobiologie*, 149: 365-386.

ZHANG, Y.; MALMQVIST, B. 1996. Relationships between labral fan morphology, body size and habitat in North Swedish blackfly larvae (Diptera: Simuliidae). *Biological Journal of the Linnean Society*, 59 (3): 261-280.

ZHANG, Y.; MALMQVIST, B. 1997. Phenotypic plasticity in a suspension-feeding insect, *Simulium lundstromi* (Diptera: Simuliidae), in response to current velocity. *Oikos*, 78:(3)503-510